

## Pancreatic Acinar Cell 3-Dimensional Culture

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**[Abstract]** Normal pancreatic acinar cells are difficult to maintain on traditional plastic culture surfaces due to their physical properties of housing large quantities of digestive enzymes and the formation of intercellular tight junctions and gap junctions (Apte and Wilson 2005; Rukstalis *et al.*, 2003). However, placing primary acinar cells within a 3-dimensional matrix (3D-culture) maintains the cells for sufficient time so that they can be monitored for physiological changes to different stimuli. We have used a modified collagen 3D-culture system that has been adapted from Means *et al.* (2005) to model the very early events associated with pancreatic cancer development. In this model, Kras<sup>G12D</sup>-expressing pancreatic acinar cells, or wildtype acinar cells treated with EGFR-dependent growth factors (*i.e.*, TGF $\alpha$ ), convert to ductal cysts that mimic the acinar-to-ductal metaplasia (ADM) stage that precedes formation of Pancreatic Intraepithelial Neoplasia (PanIN) and Pancreatic Ductal Adenocarcinoma (PDAC) (Means *et al.*, 2005; Shi *et al.*, 2013).

### Materials and Reagents

1. Rat Tail Collagen I (Life Technologies, Invitrogen™, catalog number: A10483)
2. Fetal Bovine Serum (FBS)
3. Pen-Strep
4. Hanks Balanced Salt Solution (HBSS) (Life Technologies, Gibco®, catalog number: 14065)
5. E.Z.N.Z Total RNA Kit I (Omega Bio-Tek, catalog number: R6834-01)
6. BME-phenol blue
7. Xylene
8. 10x RPMI 1640 medium (Life Technologies, Gibco®, catalog number: 31800-022) (see Recipes)
9. 4.2% NaHCO<sub>3</sub> (see Recipes)
10. Collagenase P (Roche, catalog number: 11213857001) (see Recipes)
11. 100x Soybean Trypsin Inhibitor (STI) (Sigma-Aldrich, catalog number: T6522) (see Recipes)
12. 2,000x Dexamethasone (Dex) (Sigma-Aldrich, catalog number: D4902) (see Recipes)

### 13. 3D-culture medium (see Recipes)

#### **Equipment**

1. Centrifuge (Beckman Coulter, model: GS-15R or equivalent, rotor S4180)
2. Standard 5% CO<sub>2</sub> tissue culture incubator
3. 24-well tissue culture plates
4. 37 °C shaker
5. 6 cm culture dish
6. Dissection scissors
7. 15 ml plastic screw cap tube
8. Laminar flow hood
9. Rocking platform
10. Tissue processor
11. Polypropylene Mesh (105 µm & 500 µm sizes) (Spectra/Mesh) (Fisher Scientific, catalog number: 146436 and 146418)  
*Note: Cut meshes into 10 x 10 cm squares and sterilize by autoclaving.*
12. Homogenizer spin column (Omega Bio-Tek, catalog number: HCR003)
13. Tissue-Teck Biopsy Uni-cassette (Sakura, catalog number: 4087)
14. Falcon tube (15 ml and 50 ml)
15. Nanodrop spectrophotometer

#### **Procedure**

The following procedures are for one mouse pancreas

*Notes:*

1. All procedures are performed in a laminar flow hood under sterile conditions.
  2. For each pancreas prepare 30 ml Cold HBSS + 5% FBS and 20 ml HBSS + 30% FBS.
- A. Prepare 24-well tissue culture plates (you will need 250 µl collagen mix per well)
1. On ice, mix rat tail collagen in a ratio of 0.9 ml collagen: 0.1 ml 10x RPMI 1640 medium. Add 80-100 µl 4.2% NaHCO<sub>3</sub> per 1 ml mixture (usually a light pink/yellowish color will be achieved).  
*Note: The volume of NaHCO<sub>3</sub> should be tested and adjusted for each batch of collagen.*
  2. Place each 24-well plate on ice and pipette 250 µl of collagen gel mixture into each well, assuring that the gel covers the bottom of each well. Repeat so as to have a separate plate for each day of harvest. Cover and place in a 37 °C, 5% CO<sub>2</sub> incubator for at least 1

h to solidify.

#### B. Isolate cells

3. Sacrifice each mouse following institutional IACUC protocols.
4. Put the mouse on a dissection plate in a supine position, spray with 70% EtOH and open the abdominal cavity to expose the pancreas.
5. Resect the pancreas and place into ~20 ml of HBSS in a 6 cm culture dish on ice. Swirl to wash, and decant liquid.
6. Add 5 ml cold HBSS and mince the pancreas quickly with dissection scissors.
7. Transfer the pancreas material into a 15 ml plastic screw cap tube. Add 100  $\mu$ l 10 mg/ml collagenase P to the tissue suspension and mix gently.
8. Wrap Parafilm around the cap and shake at 225 rpm in a 37 °C shaker for 15-20 min, until most of the tissue clumps are gone and the suspension look cloudy (examine by eye every 5 min to prevent over- or under-digestion).
9. Add 5 ml cold HBSS + 5% FBS and centrifuge 2,000 rpm, 2 min at 4 °C. Aspirate supernatant.
10. Resuspend the pellet 3 times in 5 ml cold HBSS + 5% FBS, spinning at 1,500 rpm, 2 min between rinses.
11. After the final wash, resuspend the pellet in 5 ml HBSS + 5% FBS.
12. With sterilized scissors, cut the tip of the 1,000  $\mu$ l pipette tip to make the opening wider and then gently pipet the cell suspension through a sterile 500  $\mu$ m mesh.
13. Pipet an additional 5 ml HBSS + 5% FBS to wash all remaining cells through the mesh.
14. Repeat by pipetting the cell suspension through a 105  $\mu$ m mesh.
15. Slowly pipet the cell suspension on top of a 50 ml tube containing 20 ml HBSS + 30% FBS.
16. Centrifuge 1,000 rpm, 2 min, 4 °C to pellet individual acini clusters.
17. Aspirate supernatant.

#### C. Plating cells

18. Resuspend the cell pellet in 8-10 ml (volume adjusted according to the size of pancreas) in 3D-culture medium.
19. Mix the collagen as above step A1.
20. Mix equal parts of the cell suspension and collagen mix.
21. Immediately plate the cell suspension in the collagen-coated wells, 0.5 ml/well (see Figure 1).

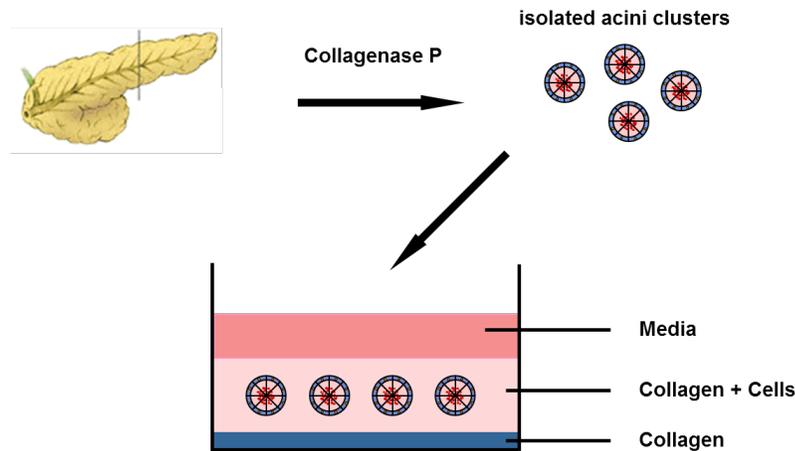


Figure 1. 3D-culture setup

22. Allow the collagen-cell mixture to solidify ~1 h at 37 °C, 5% CO<sub>2</sub>, and then add 1 ml warm 3D-culture medium (with or without growth factors inhibitors).
23. Change the medium on days 1 and 3. For *Kras*<sup>G12D</sup>-expressing acinar cells, over 90% of ductal cysts should form by day 5 (see Figure 2). Similarly, wild-type acinar cells provided 50 ng/ml TGF $\alpha$  following plating will form ductal cysts by day 5, with a conversion rate of about 70%-80% (see Figure 2) (Shi *et al.*, 2013).

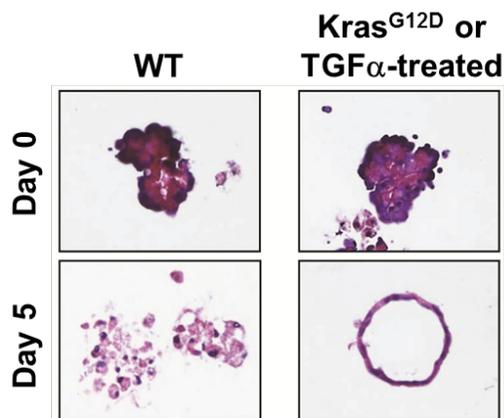


Figure 2. Pancreatic acinar cells in 3D-culture

D. RNA/protein prep from acinar cell 3D-culture

1. Dilute collagenase P (10 mg/ml) 1:50 in 1x HBSS (room temperature). Prepare 4 ml solution in a 15 ml falcon tube for each sample.
2. Wash cells/collagen disc in culture well once with 1x HBSS. Transfer collagen disc to a 15 ml falcon tube with a spatula. Pool 3 or 4 wells together to obtain enough cells.
3. Digest collagen for ~ 30 min in a 37 °C shaker, 250 rpm. Check every 10 or 15 min.

4. After all of the collagen is digested, pellet the cells at 2,000 rpm for 2 min at 4 °C.
5. Remove the supernatant, resuspend cells with 1 ml cold HBSS and transfer to a 1.5 ml centrifuge tube.
6. Centrifuge at 5,000 rpm for 2 min at 4 °C. Carefully remove the supernatant. Loosen the pellet by flicking the bottom of the tube with your finger tips.
7. OPTION 1: RNA Prep. Follow the E.Z.N.A. Total RNA Mini Kit protocol. Homogenize using a homogenizer column. Resuspend the final RNA in 40 µl ddH<sub>2</sub>O. Measure the RNA concentration using a nanodrop spectrophotometer. Store at -20 °C.
8. OPTION 2: Protein Prep. Prepare 100 µl 4x Sample Buffer (BME-phenol blue) + 1:50 protease inhibitor cocktail + 1:100 phosphatase inhibitor cocktail 1 + 1:100 phosphatase inhibitor cocktail 2 + 1:200 Na<sub>3</sub>VO<sub>4</sub> for each sample. Lyse the cells by pipetting and sonicate 15 sec. Store at -20 °C.

#### E. Collagen disc fixation

1. Fix the collagen discs in the wells with 10% neutral buffered formalin for 30 min at room temperature, and then separate the discs from the plate wall using a yellow 200 µl pipette tip and transfer the discs into scintillation vials containing 10% neutral buffered formalin. Fix on rocker at RT overnight.
2. Transfer the fixed collagen discs from the scintillation vials into standard biopsy cassettes.
3. Gradually dehydrate the collagen discs by incubating in 70%, 90%, and 100% EtOH and then in Xylenes in a glass container at room temperature, 2 times of 1 h incubation for each solution on a rocking platform.
4. After Xylene clearing, transfer the cassettes into the paraffin tank of the tissue processor. Let the collagen discs go through the last two steps of paraffin incubation (1 h each at 65 °C) and then keep them in paraffin tank until it is time for making paraffin blocks.
  - a. IMPORTANT: Keep cassettes horizontal or the gels will "flow" into the corners and will be difficult to embed.
  - b. Collagen discs can be processed through an automatic tissue processor. However, the freshness (cleanliness) of the solutions inside the processor greatly affects the outcome (the collagen discs will be hard and shrunken when solutions in the processor become old). Manual processing with small volumes of fresh solutions will keep the collagen discs in their original shape after processing.

### Recipes

1. 10x RPMI 1640 medium

Dissolve 1 package of RPMI 1640 powder in 100 ml ddH<sub>2</sub>O

- Add 2 g NaHCO<sub>3</sub>, pH to 7.2
- Filter sterilize
- Store at 4 °C
- Make fresh solution every 2 months
- 2. 4.2% NaHCO<sub>3</sub>
  - Dissolve 4.2 g NaHCO<sub>3</sub> in 100 ml ddH<sub>2</sub>O
  - Filter sterilize
  - Store at 4 °C
- 3. Collagenase P
  - Dissolve in 10 ml ddH<sub>2</sub>O to make a 10 mg/ml stock
  - Filter sterilize, aliquot and store at -80 °C
  - Thaw a new vial for each acinar prep.
- 4. 100x Soybean Trypsin Inhibitor (STI)
  - Dissolve in ddH<sub>2</sub>O to make a 10 mg/ml stock
  - Filter sterilize, aliquot and store at -20 °C
- 5. 2,000x Dexamethasone (Dex)
  - Dissolve 25 mg in 12.5 ml of 100% EtOH to generate a 2 mg/ml stock
  - Aliquot and store at -20 °C
- 6. 3D-culture medium
  - RPMI 1640
  - 1% FBS
  - 1% Pen-Strep
  - 0.1 mg/ml STI
  - 1 µg/ml Dex

### **Acknowledgments**

This protocol was adapted from Shi *et al.* (2013).

### **References**

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