

## High Resolution Detection of Genetic Changes Associated with Transposons

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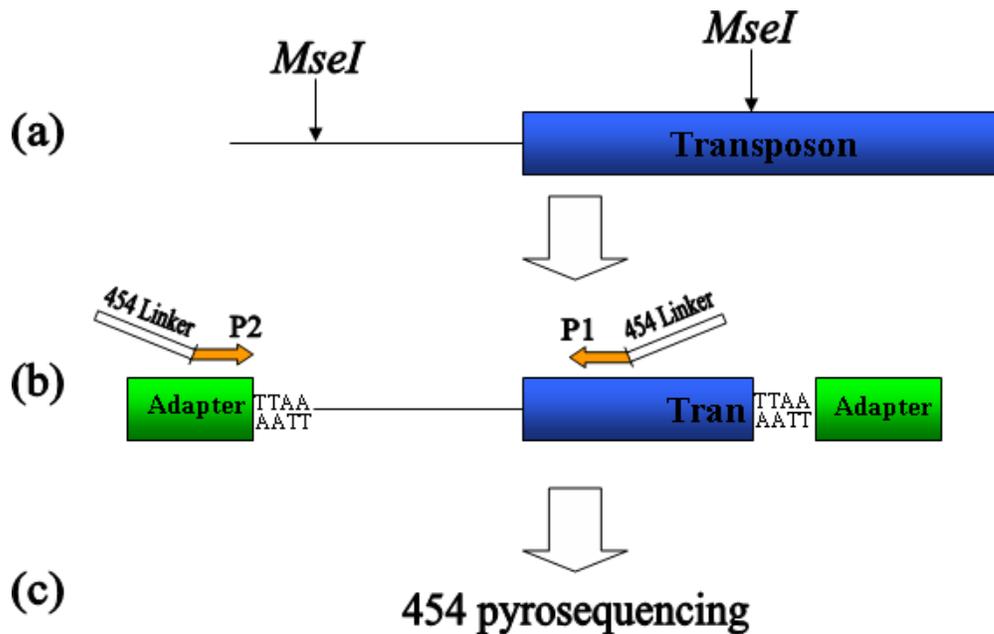
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**[Abstract]** Transposable elements (TEs) are repetitive sequences, capable of inducing genetic mutations through their transpositional activity, or by non-homologous or illegitimate recombination. Because of their similarity and often high copy numbers, examining the effects of mutations caused by TEs in different samples (tissues, individuals, species, etc.) can be difficult. Thus, high throughput methods have been developed for genotyping TEs in un-sequenced genomes. A common method is termed Transposon Display (or transposon SSAP), which utilizes restriction enzymes and PCR amplification to produce chimeric DNA molecules that include genomic and TE DNA. The advent of second generation sequencing technologies, such as 454-pyrosequencing, have dramatically improved the resolution of this assay, allowing the simultaneous sequencing of all PCR products, representing all amplified TE sites in a specific genome.

### Materials and Reagents

1. Two oligonucleotides which form the double-stranded adapter, with an overhang complementary to the overhang of the restriction enzyme used. In the case of *MseI*, the overhang is a 5' TA, and the adapter sequences are 5'- TACTCAGGACTCAT-3' and 5'- GACGATGAGTCCTGAG-3'. The two nucleotides at the 5' end of the former oligonucleotide will constitute the 5' TA overhang, after hybridization of the two sequences (green rectangles in Figure 1). These oligonucleotides should be designed such that they do not resemble known sequences in the examined species.
2. Pre-selective primers, one complementary to the adapter with the addition of a C nucleotide at the 3' end (5'-GATGAGTCCTGAGTAAC-3'; primer P2 in Figure 1), and the other complementary to the TE of interest (primer P1 in Figure 1). The TE-specific primer should be designed as a reverse-complement of the 5' end of the TE with a  $T_m=60$  °C, between 30-50 bp into the TE (to allow for sequence validation in downstream assays). Restriction enzyme recognition sites (TTAA) between the primer and the 5' end of the TE should be avoided. Both the adapter- and TE-specific primers should be designed with the linker A and B sequences (for 454-pyrosequencing) at their 5' ends.
3. NaCl
4. T4 DNA ligase and buffer (New England Biolabs, catalog number: M0202)

5. Restriction enzyme *MseI* (New England Biolabs, catalog number: R0525)
6. Taq DNA polymerase and Taq DNA polymerase buffer (EURx, catalog number: E2500)
7. MgCl<sub>2</sub>
8. dNTP mix



**Figure 1. An overview of the TD-454 pyrosequencing method.** (a) The genomic samples are fragmented with a restriction enzyme and ligated to adapters (green rectangles); (b) The fragments undergo PCR with a primer specific to the adapter (containing a 454 linker sequence) and a primer specific to the analyzed TE (containing a 454 linker sequence); (c) The resulting PCR amplicons are sequenced with a 454 pyrosequencer.

### Equipment

1. Thermal cycler

### Procedure

#### A. Adapter pair preparation

1. Mix the two adapter oligonucleotides to a final concentration of 250 ng/μl.
2. Incubate them at 95 °C for 5 min and then at room temperature (RT) for 10 min.

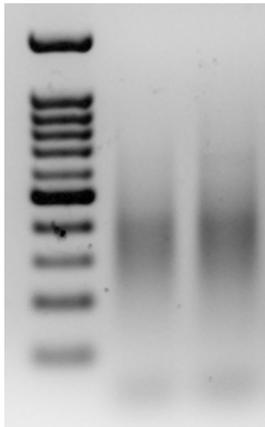
#### B. Restriction/Ligation

1. Add to a 0.2 ml tube: 1 μl of 10x ligase buffer, 1 μl of 0.5 M NaCl, 1 μl of the adapter pair,

- 120 units of T4 ligase, 2 units of *Mse*I, 300-500 ng of genomic DNA and ddH<sub>2</sub>O to a final volume of 10  $\mu$ l.
2. Mix well and incubate at 37 °C for 2-3 h.
  3. Dilute reaction 1:10 by adding 90  $\mu$ l of ddH<sub>2</sub>O.
  4. This reaction can be stored at -20 °C.

C. Pre-selective amplification

1. Add to a 0.2 ml tube: 2  $\mu$ l of 10x Taq DNA polymerase buffer, 2  $\mu$ l of 25 mM MgCl<sub>2</sub>, 0.8  $\mu$ l of dNTP mix, 1 unit of Taq DNA polymerase, 1  $\mu$ l of 50 ng/ $\mu$ l adapter-specific pre-selective primer, 1  $\mu$ l of 50 ng/ $\mu$ l transposon-specific primer, 4  $\mu$ l of Restriction/Ligation reaction products (cut with *Mse*I) and ddH<sub>2</sub>O to a final volume of 20  $\mu$ l.
2. Use the thermal cycler to PCR with the following program:
  - a. 94 °C for 3 min
  - b. 94 °C for 30 sec
  - c. 60 °C for 30 sec
  - d. 72 °C for 1 min
 Return to step b for 29 times.
3. Run the relevantly-sized PCR products (usually a narrow band between 150-550 bp, such as 300-500 bp, depending on the expected number of amplicons) in a 454-pyrosequencing machine (Figure 2).



**Figure 2.** An example of PCR products from pre-selective amplification, using a primer from the adapter and a prom from a *Stowaway*-like MITE, called *Eos*, on a 1.5% agarose gel. The lanes include a 100 bp marker (left), pre-selective amplification of goatgrass *Aegilops longissima* (accession TL05; middle lane) and pre-selective amplification of diploid wheat *Triticum urartu* (accession TMU06; right lane).

### **Acknowledgments**

The transposon (TD) display method was adapted from the amplified fragment length polymorphism (AFLP) (Vos *et al.*, 1995 Nucl Acid Res 23:4407-4414). This work was supported by a grant from the Israel Science Foundation (grant # 142/08) to Khalil Kashkush.

### **References**

1. Yaakov, B. and Kashkush, K. (2012). [Mobilization of Stowaway-like MITEs in newly formed allohexaploid wheat species.](#) *Plant Mol Biol* 80(4-5): 419-427.