

## Aldicarb-induced Paralysis Assay to Determine Defects in Synaptic Transmission in *Caenorhabditis elegans*

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**[Abstract]** Aldicarb treatment causes an accumulation of acetylcholine in the synaptic cleft of the neuromuscular junction, resulting in sustained muscle activation and eventually paralysis. Aldicarb-induced paralysis assay is an easy and fast method to determine whether synaptic transmission of a *C. elegans* mutant of interest is altered. This assay is based on the correlation of the rate of neurotransmitter release with the rate of paralysis. In this protocol, we describe a method for simultaneously assessing the aldicarb sensitivity of animals with different genotypes.

**Keywords:** Aldicarb, *C. elegans*, Acetylcholinesterase, Synaptic transmission

**[Background]** Synaptic transmission is initiated by arrival of action potential at presynaptic terminals, which in turn results in the release of neurotransmitters. The released neurotransmitters bind to and activate postsynaptic receptors (Sudhof, 2013). *C. elegans* locomotion is controlled by acetylcholine-releasing excitatory motor neurons and GABA ( $\gamma$ -aminobutyric acid)-releasing-inhibitory motor neurons (Richmond and Jorgensen, 1999; Zhen and Samuel, 2015). Acetylcholine released from cholinergic motor neurons activates acetylcholine receptors on the muscle membrane, leading to muscle excitation and contraction. Acetylcholinesterase breaks down acetylcholine in the synaptic cleft and thus terminates neurotransmission. Aldicarb is an acetylcholinesterase inhibitor. In the presence of aldicarb, acetylcholine continues to accumulate, causing persistent muscle contraction and eventual paralysis. Mutant animals with decreased levels of synaptic transmission are resistant to the paralyzing effect of aldicarb because acetylcholine more slowly accumulates in the synaptic cleft of these animals than that of wild type animals. Conversely, mutant animals with increased levels of synaptic transmission, and as a result, a faster accumulation of acetylcholine, are more sensitive to the paralyzing effect of aldicarb than wild-type animals (Rand, 2007). Thus, by comparing the time-course of aldicarb-induced paralysis, it is possible to infer the relative efficiency of synaptic transmission. However, it is necessary to note that this assay does not necessarily demonstrate that abnormal aldicarb sensitivity is a result of a presynaptic defect. For example, a defect in post-synaptic acetylcholine receptor function may also result in resistance to aldicarb (Loria *et al.*, 2004). Thus, the aldicarb-induced paralysis assay should be considered as the first step in investigating the involvement of synaptic transmission, and further corroborated by other means, such as electrophysiological analysis (Richmond, 2006).

## **Materials and Reagents**

1. 60 mm Petri dish (Genesee Scientific, catalog number: 32-105G)
2. 90% platinum, 10% iridium wire (Tritech Research, catalog number: PT-9010)
3. Copper rings (PlumbMaster, catalog number: 17668)
4. *E. coli* OP50 (University of Minnesota, Caenorhabditis Genetic Center)
5. Aldicarb (ChemService, catalog number: N-11044)
6. Ethanol (Ethyl alcohol, 190 proof, ACS-USP grade) (PHARMCO-AAPER, catalog number: 111000190)
7. Cholesterol (Sigma-Aldrich, catalog number: C3045)
8. Calcium chloride dihydrate (CaCl<sub>2</sub>·2H<sub>2</sub>O) (Sigma-Aldrich, catalog number: C3306)
9. Magnesium sulfate (MgSO<sub>4</sub>) (Sigma-Aldrich, catalog number: M7506)
10. Potassium phosphate, monobasic (KH<sub>2</sub>PO<sub>4</sub>) (Sigma-Aldrich, catalog number: P5379)
11. Potassium phosphate, dibasic (K<sub>2</sub>HPO<sub>4</sub>) (Sigma-Aldrich, catalog number: P8281)
12. Sodium chloride (NaCl) (Sigma-Aldrich, catalog number: S7653)
13. Bacto-agar (BD, Bacto™, catalog number: 214040)
14. Bacto-peptone (BD, Bacto™, catalog number: 211820)
15. 100 mM aldicarb stock solution (see Recipes)
16. 5 mg/ml cholesterol (see Recipes)
17. 1 M CaCl<sub>2</sub> stock solution (see Recipes)
18. 1 M MgSO<sub>4</sub> stock solution (see Recipes)
19. 1 M KPO<sub>4</sub>, pH 6.0 stock solution (see Recipes)
20. Nematode growth medium (NGM) agar plates (see Recipes)
21. Nematode growth medium (NGM) agar plate containing aldicarb (see Recipes)

## **Equipment**

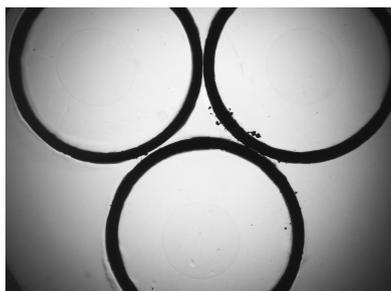
1. Forceps
2. Rotator
3. Autoclave
4. 2 L flask
5. Magnetic stir bar
6. Microscope (Leica Microsystems, model: Leica MZ6) with KL 200 LED light source (Leica Microsystems, model: KL 200 LED)

## **Software**

1. GraphPad Prism 6.0, RRID:SCR\_002798

## **Procedure**

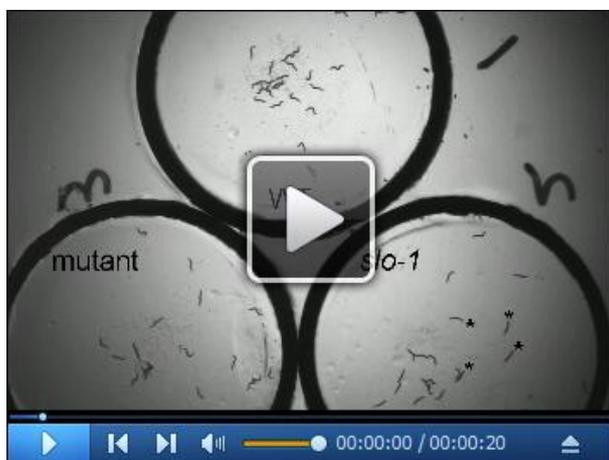
1. For the routine maintenance of worms, see Stiernagle, T. Maintenance of *C. elegans* (Stiernagle, 2006).
2. Prepare NGM agar plates containing 1 mM aldicarb (see Recipes) at least one day before the assay and store at 4 °C until use. Regular NGM agar plates for the routine maintenance should be seeded with OP50 *E. coli* and be available before the assay.
3. Pick 30-40 L4 stage wild-type N2 and mutants of interest onto separate NGM plates seeded with OP50 *E. coli* (but not containing aldicarb), and culture them at 20 °C for 20 to 24 h.
4. Use forceps to pick up a copper ring and dip into 70% ethanol. Flame the copper ring for a few seconds and immediately place it on the aldicarb-containing NGM agar plate. The copper ring will be lightly embedded in the agar surface of the plate (Figure 1). This will corral worms inside the ring. Place 3 copper rings on each assay plate and add 10 µl of OP50 *E. coli* (OD<sub>600</sub> = 1.2) at the center of the ring. Bacteria help keep the worms around the center of the ring. Let the plates dry for 30 min. Three genotypes can be assayed on the same plate.



**Figure 1. A representative image of a copper ring embedded agar plate.** Three copper rings are placed on a 60 mm NGM agar plate, and 10 µl of OP50 is added at the center of the ring.

5. Transfer 20-30 worms of each genotype from step 3 to the center of each ring. Record the number of worms placed in each ring. Stagger worm transfer every 2-3 min so that all 3 genotypes are examined within a 10-min interval.
6. Observe the worms in a 10-min interval. If there are worms not moving, then gently prod their heads with a platinum wire until they move (Videos 1 and 2). If they fail to move their head in response to even harsh touch, then remove them from the plate and record. Continue until no worm is left. Make sure that all of the worms are accounted for throughout the assay.

**Video 1. A video recording of responses of worms with different genotypes in the presence of aldicarb.** The video (120 frames) was recorded for 4 min after 50 min incubation on an aldicarb plate. \* indicates the paralyzed worms. WT: wild-type animals, *slo-1*: *slo-1(eg142lf)*, mutant: an uncharacterized mutant.

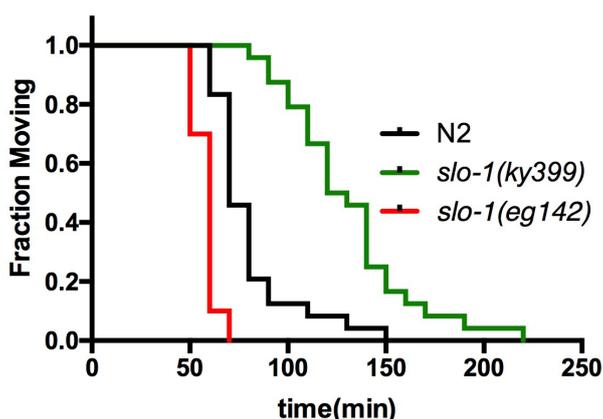


**Video 2. A video recording shows paralyzed worms that do not respond to harsh touch**



### Data analysis

1. Software: Prism 6.0
2. Use 'Survival analysis' function to plot the survival graph (Figure 2). At least 20 worms of each genotype are used per assay. Repeat the assay two more times on different days (total 3 trials) and report the summary of the statistics as shown in Tables 1A and 1B.



**Figure 2. Survival curve for aldicarb-induced paralysis assay.** Wild-type, *slo-1(ky399 gain-of-function)* and *slo-1(eg142 loss-of-function)* mutant animals were simultaneously tested for aldicarb sensitivity in a copper ring-embedded assay plate (adapted from Oh *et al.*, 2017).

**Table 1A. Median survival time of *C. elegans* exposed to aldicarb** (Median survival unit is minutes. The number in parenthesis is sample number)

	Trial 1	Trial 2	Trial 3
N2	70(24)	80(30)	70(29)
<i>slo-1(ky399)</i>	125(24)	110(23)	90(23)
<i>slo-1(eg142)</i>	60(20)	60(27)	50(26)

**Table 1B. P values of Log-rank test**

	Trial 1	Trial 2	Trial 3
N2 vs. <i>slo-1(ky399)</i>	< 0.0001	< 0.0001	< 0.0001
N2 vs. <i>slo-1(eg142)</i>	< 0.0001	< 0.0001	< 0.0001
<i>slo-1(ky399)</i> vs. <i>slo-1(eg142)</i>	< 0.0001	< 0.0001	< 0.0001

3. Statistical analysis: The log-rank test is used to analyze the data. Prism software returns overall curve comparison results. However, pair-wise comparisons can be performed to compare 2 specific groups. Statistically significant *P*-values need to be adjusted for multiple comparisons if more than 2 pair-wise comparisons are performed. A Bonferroni corrected threshold is used to determine statistically significant *P*-values ([http://www.graphpad.com/guides/prism/6/statistics/index.htm?survival\\_curves.htm](http://www.graphpad.com/guides/prism/6/statistics/index.htm?survival_curves.htm)). In this example, 3 comparisons, N2 vs. *slo-1(ky399)*, N2 vs. *slo-1(eg142)*, and *slo-1(ky399)* vs. *slo-1(eg142)*, are performed and *P*-values for each comparison is obtained. If threshold for statistical significance is set at a *P*-value of 0.05, then any *P*-value that is less than the corrected threshold, *i.e.*, 0.05 divided by 3, is considered statistically significant in this example.

## **Notes**

1. As per standard scientific procedure, the experimenter should be blinded to the genotypes of the test worms. Ideally a colleague should place the worms (procedure step 5) and label the worms to prevent the experimenter from identifying the genotypes. For mutants with obvious phenotypes, an additional mutant with a similar phenotype, which is unrelated to the study of interest, may be added.
2. The resolution of differential sensitivity can be changed by varying the concentration of aldicarb. For example, mutants that have higher rates of neurotransmission than wild type worms may be better distinguishable at a lower concentration of aldicarb (e.g., 0.5 mM), which increases the time for the worms to get paralyzed.

## **Recipes**

1. 100 mM aldicarb stock solution  
 Dissolve 100 mg of aldicarb in 5.25 ml of 70% ethanol at room temperature  
 Store at 4 °C  
*Note: We have stored the stock up to 2 weeks and have not tried longer storage.*
2. 5 mg/ml cholesterol  
 Add 250 mg of cholesterol in 50 ml of 95 % ethanol and mix by rotating on a rotator at room temperature. It takes a few hours to dissolve. Store at room temperature
3. 1 M CaCl<sub>2</sub> stock solution  
 Dissolve 14.7 g of CaCl<sub>2</sub>·2H<sub>2</sub>O in 100 ml ddH<sub>2</sub>O and autoclave for 30 min at 121 °C. Store at room temperature
4. 1 M MgSO<sub>4</sub> stock solution  
 Dissolve 12.04 g of MgSO<sub>4</sub> in 100 ml ddH<sub>2</sub>O and autoclave for 30 min at 121 °C. Store at room temperature
5. 1 M KPO<sub>4</sub>, pH 6.0 stock solution  
 Make 500 ml of 1 M KH<sub>2</sub>PO<sub>4</sub> (monobasic) and 250 ml of 1 M K<sub>2</sub>HPO<sub>4</sub> (dibasic) solution. While measuring pH of 1 M KH<sub>2</sub>PO<sub>4</sub> (pH is below 5), add and stir 1 M K<sub>2</sub>HPO<sub>4</sub> slowly until pH reaches 6.0. It will take less than 250 ml of K<sub>2</sub>HPO<sub>4</sub> to reach pH 6.0. Aliquot the solution and autoclave for 30 min at 121 °C. Store at room temperature
6. Nematode growth medium (NGM) agar plate
  - a. Add 3 g of NaCl, 16 g of Bacto-agar, and 2.5 g of Bacto-peptone in a 1 L of ddH<sub>2</sub>O in a 2 L flask with magnetic stir bar
  - b. Autoclave for 30 min at 121 °C
  - c. Let the NGM agar cool to 55 to 60 °C while stirring on a stirrer, Add 1 ml of 5 mg/ml cholesterol, 1 ml of 1 M CaCl<sub>2</sub>, 1 ml of 1 M MgSO<sub>4</sub>, and 25 ml of 1 M KPO<sub>4</sub>, pH 6.0 while stirring

- d. Dispense 10 ml per 60 mm Petri dish and let them solidify without perturbation
7. Nematode growth medium (NGM) agar plate containing aldicarb
  - a. For aldicarb-containing NGM plates, add aldicarb solution to a desired final concentration along with cholesterol, CaCl<sub>2</sub>, MgSO<sub>4</sub>, and 1 M KPO<sub>4</sub>, pH 6.0 when NGM agar medium is cooled to 55 to 60 °C
  - b. For 1 mM aldicarb, add 1 ml of 100 mM aldicarb for 100 ml of NGM. Pour 10 ml per 60 mm Petri dish and let them dry for at least 1 day. Plates can be stored at 4 °C

*Note: We have used plates stored at 4 °C for up to 2 weeks without any issue.*

### **Acknowledgments**

The protocol has been adapted from Oh *et al.* (2017), eLife 6: e24733. This work was supported, in part, by a grant from the National Institute of Health.

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